

# ichrom∝™ NORO

# INTENDED USE

Infection

ichroma™ NORO is a fluorescence Immunoassay (FIA) for the qualitative determination of norovirus in <u>human feces</u>. It is useful as an aid in screening of norovirus infection.

For in vitro diagnostic use only.

# INTRODUCTION

Noroviruses are the most common cause of epidemic gastroenteritis, accounting for more than 23 million cases of gastroenteritis annually in the United States and about 50% of all cases of outbreaks worldwide, and they are a significant cause of sporadic cases of community-related gastroenteritis<sup>11</sup>. Noroviruses, which belong to the family Caliciviridae, are small non-enveloped RNA viruses that possess a linear, positive-sense, and single-stranded RNA genome. Noroviruses are genetically classified into 5 groups (GI V); GI, GII, and GIV cause human infections<sup>21</sup>. The most common disease, norovirus gastroenteritis often presents with nausea, vomiting, and watery diarrhea, following an incubation period of 1–2 days. Other associated symptoms include abdominal pain or cramps, anorexia, malaise, and low-grade fever<sup>31</sup>. This leads to an estimated 50,000 child deaths every year, nearly all of which occur in developing countries<sup>41</sup>.

Diagnosis may be made by rapid detection of norovirus antigen in stool specimens. The "ichroma" NORO" is an immunoassay for the detection of GI, GII human norovirus in stool sample.

#### PRINCIPLE

The test uses a sandwich immunodetection method; Dried detector antibody in detection buffer, once diluted with the diluent, bind with antigen in the sample to form antibody-antigen-antibody complexes. These complexes then migrates through the nitrocellulose matrix and are captured by another sets of immobilized antibodies on the test line.

The more antigens in the sample, the more antigen-antibody complexes, which leads to stronger fluorescence signal. This signal then is interpreted by the reader to display 'norovirus positive' in sample.

#### COMPONENTS

ichroma<sup>™</sup> NORO consists of 'Cartridges', 'Detection Buffer Tubes', 'Diluent vial', 'Sample collection tubes', 'Filter caps', 'Sample swabs', 'ID chip' and an 'Instruction for use'.

- The cartridge contains a test strip, the membrane which has norovirus antibody at the test line, with chicken IgY at the control line.
- Each cartridge is individually sealed in an aluminum foil pouch containing a desiccant. 25 sealed cartridges are packed in a box which also contains an ID chip.
- The detection buffer contains norovirus antibody fluorescence conjugate, anti-chicken IgY fluorescence conjugate, bovine serum albumin (BSA) as a stabilizer and sodium azide as a preservative in phosphate buffered saline (PBS).
- The dried detection buffer is pre-dispensed in a tube. 25 detection buffer tubes are packed in an aluminum foil pouch.
- The diluent contains a detergent and bovine serum albumin (BSA) as a stabilizer and sodium azide as a preservative in sodium borate buffer. The diluent is dispensed in a vial.
- The sample collection tube contains sample dilution buffer. It contains bovine serum albumin (BSA), tween20, sodium chloride

and sodium azide in Tris-HCl and DDW.

 25 sample collection tubes are packaged in a box and further packed in a Styrofoam box with ice-pack for the shipment.

#### WARNINGS AND PRECAUTIONS

- For in vitro diagnostic use only.
- Carefully follow the instructions and procedures described in this 'Instruction for use'.
- Use only fresh samples and avoid direct sunlight.
- Lot numbers of all the test components (cartridge, ID chip, detection buffer, and diluent sample collection tube) should agree.
- Do not interchange test components between different lots or use test components after the expiration date, either of which might yield misleading of test result(s).
- Do not reuse. A detection buffer tube should be used for processing one sample only. So should a cartridge.
- The cartridge should remain sealed in its original pouch before use. Do not use the cartridge, if is damaged or already opened.
- Frozen sample should be thawed only once. For shipping, samples must be packed in accordance with applicable local requirement.
- Just before use, allow the cartridge, the detection buffer and reach the room temperature by leaving them in the room for approximately 30 minutes at the least.
- ichroma<sup>™</sup> NORO as well as the instrument for ichroma<sup>™</sup> tests should be used away from vibration and/or magnetic field. During normal usage, it can be noted that instrument for ichroma<sup>™</sup> tests may produce minor vibration.
- Used detection buffer tubes, pipette tips, cartridges, sample collection tubes, filter caps and sample swab should be handled carefully and discarded by an appropriate method in accordance with relevant local regulations.
- An exposure to larger quantities of sodium azide may cause certain health issues like convulsions, low blood pressure and heart rate, loss of consciousness, lung injury and respiratory failure.
- ichroma<sup>™</sup> NORO will provide accurate and reliable results subject to the following conditions.
- Use ichroma<sup>™</sup> NORO should be used only in conjunction with the instrument for ichroma<sup>™</sup> tests.

# STORAGE AND STABILITY

- The cartridge is stable for 20 months (while sealed in an aluminum foil pouch) if stored at 4-30 °C.
- The detection buffer, the diluent and the sample collection tube are stable for 20 months if stored at 2-8 °C.
- After the cartridge pouch is opened, the test should be performed immediately.

#### LIMITATION OF THE TEST SYSTEM

- The test may yield false positive result(s) due to the crossreactions and/or non-specific adhesion of certain sample components to the capture/detector antigens.
- The test may yield false negative result. The non-responsiveness
  of the antibodies to the antigens is most common where the
  epitope is masked by some unknown components, so as not to
  be detected or captured by the antibodies. The instability or
  degradation of the antigen with time and/or temperature may
  cause the false negative as it makes antibody unrecognizable by
  the antigens.
- Other factors may interfere with the test and cause erroneous results, such as technical/procedural errors, degradation of the test components/reagents or presence of interfering substances in the test samples.
- Any clinical diagnosis based on the test result must be supported by a comprehensive judgment of the concerned physician including clinical symptoms and other relevant test results.

# MATERIALS SUPPLIED

# REF CFPC-76

#### Components of ichroma<sup>™</sup> NORO

Box Contains:	
- Cartridges	25
- Detection buffers	25
<ul> <li>Sample collection tubes</li> </ul>	25
- Filter caps	25
- Sample Swabs	25
- Diluent vial	1
- ID chip	1
- Insert	1

#### MATERIALS REQUIRED BUT SUPPLIED ON DEMAND

Following items can be purchased separately from ichroma<sup>™</sup> NORO. Please contact our sales division for more information.

- ichroma™ II REF FPRR021
- Boditech NORO Control REF CFPO-165

# SAMPLE COLLECTION AND PROCESSING

The sample type for ichroma™ NORO is human feces.

- Fecal samples must be taken as soon as the symptoms (diarrhea) appear.
- Collect random samples of feces in a clean, dry container or a receptacle, making sure to exclude.
- Loosen a cap on the upper part of a sample collection tubes and remove. Use with
- care not to spill or splatter solution from the tube.
- Collect random samples by using a sample swab with the proper method; insert the
- sample swab and put the stick into the fecal samples several times (5-6 times) at different sites so as to get a representative sampling.
- ※ Fill up the groove of a sample swab with fecal samples and please check whether the quantity is too much or not.
- Insert the sample swab into the sample collection tube.
- Swirl the sample swab at least 10 times. And break the sample swab at the break
- point and remove top portion of swab.
- Assemble filter cap on the sample collection tube.
- ★ Mixture may be stored for up to 3 days at 2-8 °C in a darkroom

#### TEST SETUP

- Check the contents of ichroma<sup>™</sup> NORO: Sealed Cartridges, Detection Buffer Tubes, Diluent Vial, 'Sample collection tubes', 'Filter caps', 'Swabs' and ID Chip.
- Ensure that the lot number of the cartridge matches that of the ID chip, the detection buffer as well as the diluent.
- Keep the sealed cartridge (if stored in refrigerator), the detection buffer tube and the diluent at room temperature for at least 30 minutes just prior to the test. Place the cartridge on a clean, dustfree and flat surface.
- Turn on the Instrument for ichroma<sup>™</sup> tests.
- Insert the ID Chip into the ID chip port of the Instrument for ichroma™ tests.
- Press the 'Select' button on the Instrument for ichroma™ tests. (Please refer to the 'Instrument for ichroma™ tests Operation Manual' for complete information and operating instructions.)



#### TEST PROCEDURE



- Put a sample swab into the fecal sample about 5-6 times at different sites and try to avoid obtaining clumps of feces.
- Loosen a cap on the upper part of a sample collection tube and remove.
- 3) Insert the sample swab into the sample collection tube.
- Swirl the sample swab at least 10 times. And break the sample swab at the break point and remove top portion of sample swab.
- Assemble filter cap on the sample collection tube and the filter cap end.
- 6) Break the break point of filter cap.
- Open the diluent vial and transfer 150 μL of diluent using a pipette to the detection buffer tube.
- Transfer only 2 drops (about 30 μL) of feces sample using a sample collection tube to the Detection buffer tube.
- Mix well by pipetting 10-20 times. (The sample mixture must be used immediately.
- Pipette out 100 µL of a sample mixture and dispense it into the DB well on the cartridge.
- 11) Leave the sample-loaded cartridge at room temperature for 12 minutes

▲ <u>Scan the sample-loaded cartridge immediately when the</u>

incubation time is over. If not, it will cause inexact test result.

- 12) To scan the sample-loaded cartridge, insert it into the cartridge holder of the instrument for ichroma<sup>TM</sup> tests. Ensure proper orientation of the cartridge before pushing it all the way inside the cartridge holder. An arrow has been marked on the cartridge especially for this purpose.
- 13) Tab the 'Start' icon on the screen.
- Instrument for ichroma™ tests will start scanning the sampleloaded cartridge immediately.
- Read the test result on the display screen of the instrument for ichroma™ tests.

#### INTERPRETATION OF TEST RESULT

 Instrument for ichroma<sup>™</sup> tests calculates the test result automatically and displays "Positive / Negative / Indeterminate".

<ul> <li>Ancinary</li> </ul>	<ul> <li>Anclinary value is served in the form of a cut-on index (COI).</li> </ul>		
Cut-off index (COI)	Result	Note	
≤ 0.9	Negative for norovirus	No need to additional test.	
> 0.9, < 1.0	Indeterminate	Need to retest.	
≥ 1.0	Positive norovirus	Need to confirmation test.	

# QUALITY CONTROL

- Quality control tests are a part of the good testing practice to confirm the expected results and validity of the assay and should be performed at regular intervals.
- The control tests should be performed immediately after opening a new test lot to ensure the test performance is not altered.
- Quality control tests should also be performed whenever there is any question concerning the validity of the test results.
- Control materials are not provided with ichroma<sup>™</sup> NORO. For more information regarding obtaining the control materials, contact Boditech Med Inc.'s Sales Division for assistance. (Please refer to the instruction for use of control material.)

### PERFORMANCE CHARACTERISTICS

#### Analytical Sensitivity

Two Norovirus genogroup samples (genotype GI-4, GII\_3,4,6,16,17) were tested with the **ichroma™ NORO**. All of genotyped samples were shown positive

Commercial ELISA		ichrom	ichroma <sup>™</sup> NORO	
Dilution factor	Result	COI value	Result	
Original	Positive	50.02	Positive	
1/5	Positive	41.13	Positive	
1/10	Positive	28.90	Positive	
1/50	Positive	11.74	Positive	
1/100	Positive	5.08	Positive	
1/200	Positive	2.74	Positive	
1/400	Negative	1.63	Positive	
1/800	Negative	1.03	Positive	
1/1600	Negative	0.75	Negative	
1/3200	Negative	0.33	Negative	

### Analytical Specificity

#### - Cross-reactivity

There was no false positive result from 17 species virus samples and 24 species bacteria samples containing potentially interfering substances with the **ichroma™ NORO** test.

The overall	specificity	was	100 %.	
				-

		Virus	
#1	ADV1	#10	Rotavirus (DS-1)
#2	ADV2	#11	enterovirus type 71
#3	ADV3	#12	cytomegalovirus
#4	ADV4	#13	poliovirus type 1
#5	ADV5	#14	Coxsackie virus B type 5
#6	ADV31	#15	Coxsackie virus B type 6
#7	ADV40	#16	herpes simplex virus type 1
#8	ADV41	#17	herpes simplex virus type2
#9	Rotavirus (WA)		
		Bacteri	a
#1	Staphylococcus	aureus (ATC	C 29213)
#2	Enterococcus fa	aecalis (ATCC	29212)
#3	Escherichia col	i (ATCC 25922	2)
#4	Kleb-siella oxyt	oca (ATCC 70	0432)
#5	#5 Pseudomonas aeruginosa (ATCC 27853)		
#6	#6 Neisseria gonorrheae (ATCC 27853)		
#7	Aeromonas hydrophila (KCCM 32586)		
#8	Enterobacter cloacae (KCCM 32586)		
#9	Vibrio parahaemolyticus (KCCM11965)		
#10	Salmonella group B (Clinical isolate from patient)		
#11	Salmonella group C (Clinical isolate from patient)		
#12	2 Salmonella group D (Clinical isolate from patient)		
#13	\$13 Salmonella group E(Clinical isolate from patient)		
#14	\$14 Shigella group D (Clinical isolate from patient)		
#15	#15 Staphylococcus epidermidis (Clinical isolate from patient)		
#16	#16 Serratia marcescens (Clinical isolate from patient)		
#17	#17 Yersinia enterocolitica (Clinical isolate from patient)		
#18	#18 Salmonella typhi (Clinical isolate from patient)		
#19	Clostridium dif	ficile (Clinical	isolate from patient)
#20	Candida albica	ns (Clinical iso	plate from patient)
#21	Candida paraps	silosis (Clinica	al isolate from patient)
#22	Campylobacter	spp	
#23	Proteus vulgari	s	

# bditech

## #24 Proteus mirabilis

#### Interference

There, in test samples, are biomolecules and chemical drugs were added to the test samples at concentrations much higher than their normal physiological levels in human feces.

ichroma<sup>™</sup> NORO test results did not show any significant interference with these biomolecules and chemical drugs

Biomolecule			
#1	Bilirubin	#4	Cholesterol
#2	Hemoglobin	#5	BSA
#3	Triglycerides		
	Chemical	l drug	
#1	cephradine	#9	metronidazole
#2	cefuroxime axetil	#10	ibuprofen
#3	cefpodoxime proxetil	#11	acetaminophen
#4	cefixime	#12	barium sulfate
#5	tetracycline hcl	#13	DMSO
#6	levofloxacin	#14	DMF
#7	amoxicillin	#15	DDW
#8	loperamide oxide	#16	PBS

Precision

#### - Between lot

One person tested three different lots of ichroma<sup>m</sup> NORO, ten times at each concentration of the control standard.

- Between person

Three different persons tested one lot of **ichroma™ NORO**, Five times at each concentration of the control standard.

- Between day

One person tested one lot of **ichroma™ NORO** during three days, Five times at each concentration of the control standard.

#### - Between site

One person tested **ichroma™ NORO** at three different site, five times at each concentration of the control standard.

	Between LC	Ts	Between per	rson
Cal.	Positive /Sample number	Positive	Positive /Sample number	Positive
Negative	0/10	0 %	0/5	0 %
Low	10/10	100 %	5/5	100 %
Mid	10/10	100 %	5/5	100 %
	Between da	ау	Between si	ite
Cal.	Positive /Sample number	Positive	Positive /Sample number	Positive
Negative	0/10	0 %	0/5	0 %
Low	10/10	100 %	5/5	100 %
Mid	10/10	100 %	5/5	100 %

#### Comparability with reference product

	Reference NORO assay			
		Positive	Negative	Total
	Positive	78	1	79
ichroma™ NORO	Negative	10	99	109
Nono	Total	88	100	188

- Clinical sensitivity = 88.6 %

- Clinical specificity = 99.0 %

#### REFERENCES

- Kim et al., Evaluation of the SD Bioline Norovirus rapid immunochromatography test using fecal specimens from Korean gastroenteritis patients., JVM. 186 (2012) 94–98., 2012.
- Park KS et al., Evaluation of a New Immunochromatographic Assay Kit for the Rapid Detection of Norovirus in Fecal Specimens. Ann Lab Med., Jan;32(1):79-81., 2012.
- Hoonmo L.K et al., Noroviruses: The Principal Cause of Foodborne Disease Worldwide., NIH Public Access., 10(50): 61– 70., 2010.
- 4. CDC/Norovirus: http://www.cdc.gov/norovirus/worldwide.html

Note: Please refer to the table below to identify various symbols.

Σ	Sufficient for <n> tests</n>
(ÌI	Read instruction for use
$\Box$	Use by Date
LOT	Batch code
REF	Catalog number
$\wedge$	Caution
-	Manufacturer
EC REP	Authorized representative of the European Community
IVD	In vitro diagnostic medical device
X	Temperature limit
8	Do not reuse
CE	This product fulfills the requirements of the Directive 98/79/EC on in vitro diagnostic medical devices

For technical assistance; please contact:

Boditech Med Inc.'s Technical ServicesTel:+82 33 243-1400

sales@boditech.co.kr

E-mail:

# Boditech Med Incorporated

43, Geodudanji 1-gil, Dongnae-myeon, Chuncheon-si, Gang-won-do, 24398 Republic of Korea Tel: +(82) -33-243-1400 Fax: +(82) -33-243-9373 www.boditech.co.kr

# EC REP Obelis s.a

Bd. Général Wahis 53, 1030 Brussels, BELGIUM Tel: +(32) -2-732-59-54 Fax: +(32) -2-732-60-03 E-Mail: mail@obelis.net

CE

